

# THE EFFECTS OF FRUCTOOLIGOSACCHARIDE AND GALACTOOLIGOSACCHARIDE SUPPLEMENTATION ON IMMUNE ACTIVITY IN WISTAR RATS

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## ABSTRACT

Currently, there is controversy over the potential benefits of fermentable carbohydrate (prebiotic) supplements presently being added to infant formula. Two of the most commonly used prebiotic supplements are fructooligosaccharides (FOS) and galactooligosaccharides (GOS). It has been proposed that when these two prebiotics are added to formula, they promote the growth of bifidobacteria and lactobacilli in the infant gut, ultimately fostering a microbial and immunological environment similar to that which results from ingesting human milk oligosaccharides. In order to determine whether these dietary interventions result in a permanent impact on the immune system, two cohorts of Wistar rats each ingested either FOS, GOS or a control diet for 10 days in early infancy. Systemic and mucosal immune tissues were collected from these rats at 16 or 70 days of age and were subsequently examined to compare cytokine and chemokine concentrations, key markers of immunological activity. The findings resulting from this study indicated differences in immune activity between sexes, but did not demonstrate a benefit of FOS or GOS feeding during infancy on the immune system.

## INTRODUCTION

- Breastfed infants possess a number of health advantages relative to formula fed infants<sup>1</sup>.
- Human milk oligosaccharides (HMOs), which act as prebiotics<sup>1</sup>, appear to foster healthy gut and immune system development<sup>2</sup>.
- FOS and GOS are potential HMO supplements, currently being added to infant formula<sup>3</sup>, however, the benefits of these prebiotics are widely debated.
- Recent research has also suggested that there are sex-based differences in the processing of prebiotics, such as FOS<sup>10</sup>.
- By examining the immunological responses during both acute and prolonged periods of FOS and GOS supplementation, it is possible to determine whether or not the immune system is impacted by dietary intervention during development.

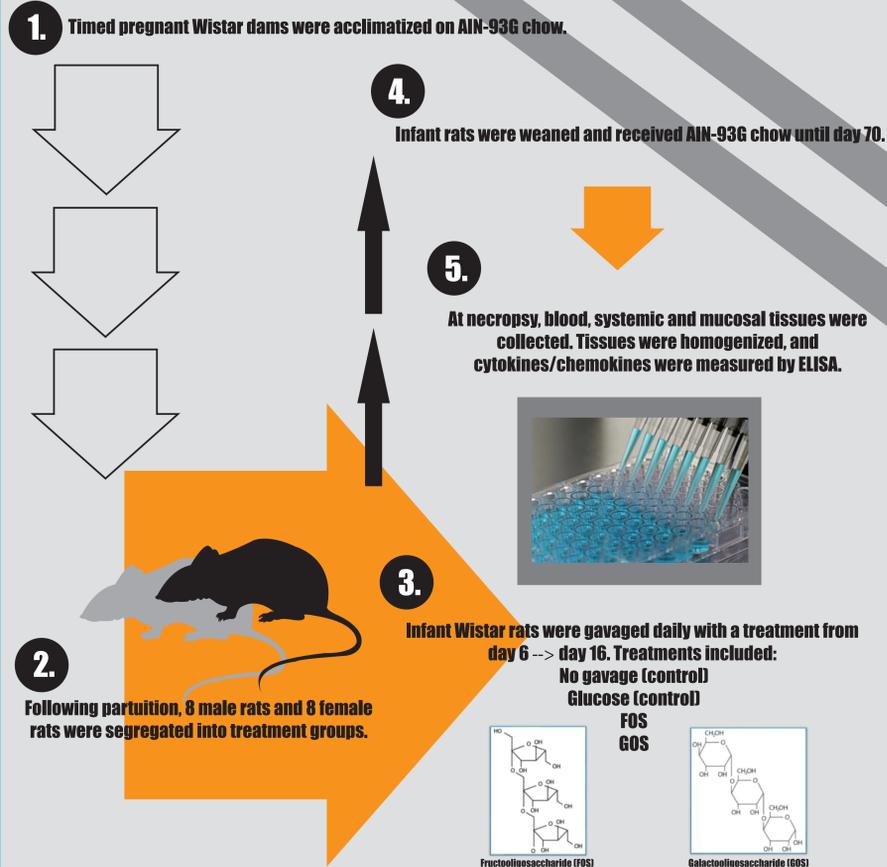
### Proposed Benefits of FOS & GOS Ingestion:

- Stimulates bifidobacterial growth<sup>4</sup>
- Immune benefits<sup>10</sup>
- Increased faecal SIgA concentrations<sup>2,12</sup>
- Increased salivary SIgA<sup>12</sup>
- Resistance to allergy development<sup>1,5</sup>
- Lowering intestinal pH<sup>13</sup>

### Proposed Dangers of FOS & GOS Ingestion:

- Increased intestinal permeability, resulting in increased bacterial translocation<sup>1</sup>
- Increased systemic endotoxin presence<sup>7</sup>
- Increase in proinflammatory markers<sup>15</sup>
- Increased SIgA in the GI tract<sup>8</sup>

## METHODS



## RESULTS

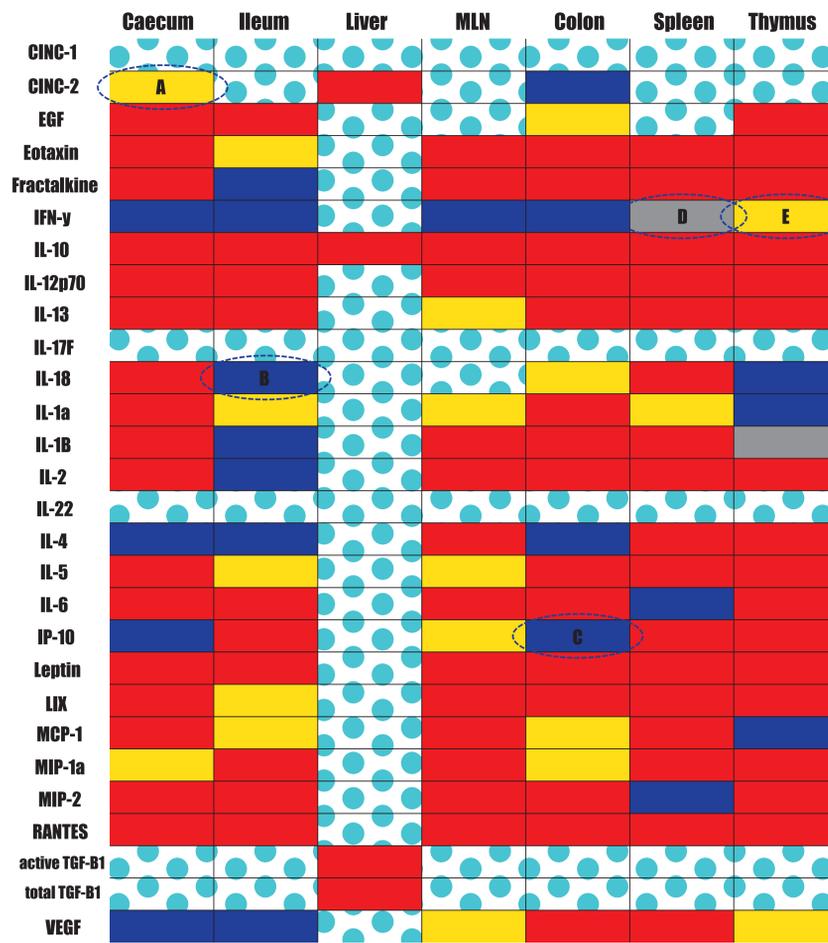


Figure 1. Cytokine/chemokine concentrations measured from tissue of 16-day-old rats gavaged with either a glucose control, FOS, GOS, or a no gavage control for 10 days during infancy. Significant differences between treatment groups and sex were determined using Tukey's multiple comparison test.

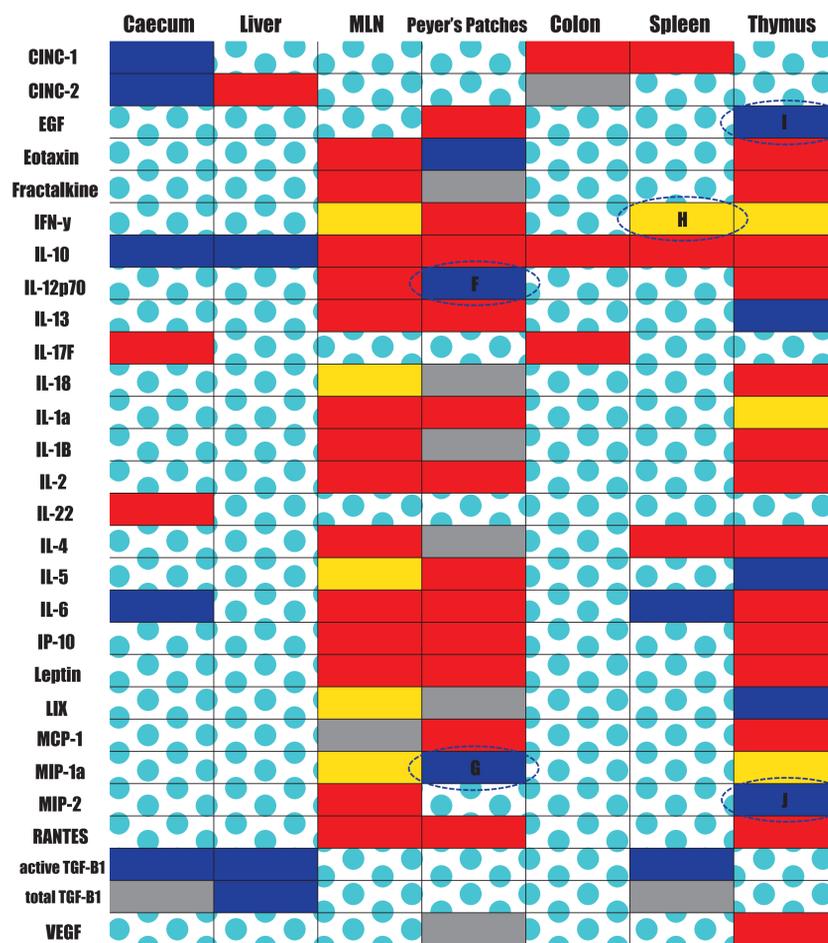
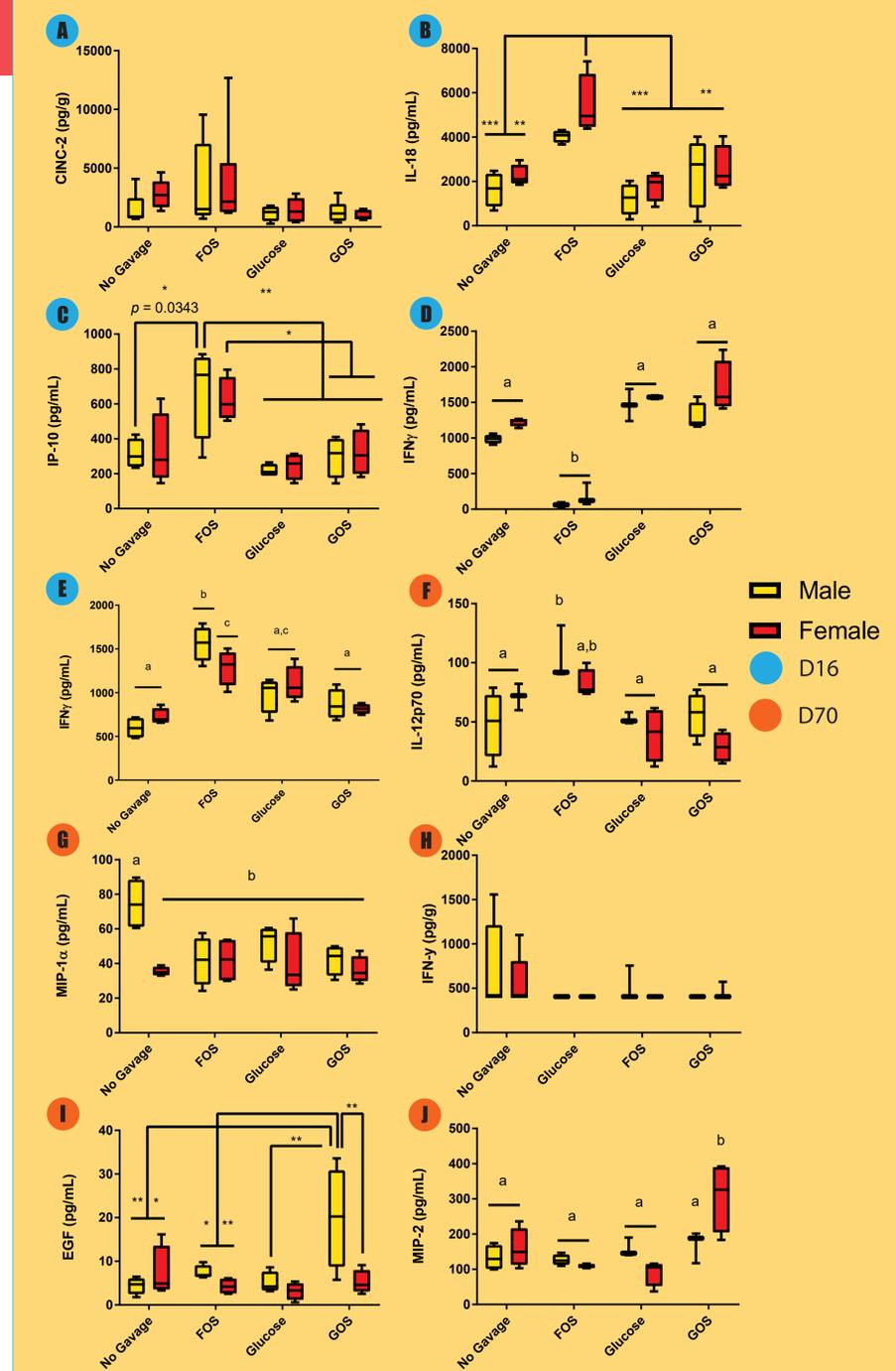


Figure 2. Cytokine/chemokine concentrations measured from tissue of 70-day-old rats gavaged with either a glucose control, FOS, GOS, or a no gavage control for 10 days during infancy. Significant differences between treatment groups and sex were determined using Tukey's multiple comparison test.



## CONCLUSIONS

- Differential impacts of diet and sex on cytokine and chemokine profiles were observed mucosally and systemically.
- FOS- and GOS- supplemented diets had minimal impact on cytokine profiles in the tissues investigated from day 16 rats, while tissues from day 70 rats illustrated significant differences primarily linked to sex rather than diet, a finding in keeping with our earlier studies of FOS fed rats<sup>10</sup>.
- Overall:** changes in immune activity varied between systemic and mucosal tissue, suggesting that sex-based differences account for more effects than any dietary intervention tested.

## REFERENCES

- Abrahamsson, T. R., Jakobsson, H. E., Andersson, A. F., Björkstén, B., Engstrand, L., and Jenmalm, M. C. (2014). Low gut microbiota diversity in early infancy precedes asthma at school age. *Clinical & Experimental Allergy*, 44(6):842-850. Doi: 10.1111/cea.12253.
- Bakker-Zierikzee, A. M., van Tol, E. A. F., Kroes, H., Alles, M. S., Kok, F. J., and Bindels, J. G. (2006). Faecal SIgA secretion in infants fed on pre- or probiotic infant formula. *Paediatric Allergy and Immunology*, 17:134-140.
- Barrat, E., Michel, C., Poupeau, G., David-Sochoard, A., Rival, M., et al. (2008). Supplementation with galactooligosaccharides and inulin increases bacterial translocation in artificially reared newborn rats. *Pediatric Research*, 64(1):34-39.
- Bode, L. (2012). Human milk oligosaccharides: every baby needs a sugar mama. *Glycobiology*, 22(9):1147-1162. Doi: 10.1093/glycob/cw8074.
- Bouchard, G., Castan, L., Chesne, J., Braza, F., Aubert, P., Neunlist, M., Maggian, A., and Bodinier, M. (2016). Maternal exposure to GOS/inulin mixture prevents food allergies and promotes tolerance in offspring in mice. *European Journal of Allergy and Clinical Immunology*, 7:168-76.
- Chierici, R., Faniara, S., Saccomandi, D., and Vigi, V. (2003). Advances in the modulation of the microbial ecology of the gut in early infancy. *Acta paediatrica*, 44:156-63.
- Clarke, S. T., Green-Johnson, J. M., Brooks, S. P. J., Ramdath, D. D., Bereik, P., Avila, C., Inglis, B., Yanke, L. J., Selinger, B., and Kalmokoff, M. (2016). D2-1 Fructan supplementation alters host immune responses in a manner consistent with increased exposure to microbial components: results from a double-blinded, randomised, cross-over study in healthy adults. *British Journal of Nutrition*, 115(1):78-1750. Doi: 10.1017/S0007414516000908.
- Genda, T., Sasaki, Y., Koyudo, T., Hino, S., Nishimura, N., Tsukahara, T., Sonoyama, K., and Morita, T. (2017). Fructooligosaccharide-induced transient increases in fecal immunoglobulin A concentrations in rats are associated with mucosal inflammation in response to increased gut permeability. *The Journal of Nutrition*, 147(10):1900-1908. Doi: 10.3945/jn.117.253055.
- Lathrop, S. K., Bloom, S. M., Rao, S. M., Nutsch, K., Liu, C. W., Santacruz, N., Peterson, D. A., Stappenbeck, T. S., and Hsieh, C. S. (2011). Peripheral education of the immune system by colonic commensal microbiota. *Nature*, 478(7368):250-254. Doi: 10.1038/nature10334.
- Moro, G., Arslanoglu, S., Stahl, B., Jelenc, J., Wahn, U., and Boehm, G. (2006). A mixture of prebiotic oligosaccharides reduces the incidence of atopic dermatitis during the first six months of age. *Archives of Disease in Childhood*, 91(10):814-819. Doi: 10.1136/adc.2006.098251.
- Oliveira, D. L., Wilbey, R. A., Grandison, A. S., and Roseiro, L. B. (2015). Milk oligosaccharides: a review. *International Journal of Dairy Technology*, 68(3):305-321. Doi: 10.1111/1471-0307.12209.
- Scholten, P. A. M. J., Alliet, P., Raes, M., Alles, M. S., Kroes, H., Boehm, G., Knippels, L. M. J., Knol, J., and Vandenplas, Y. (2008). Fecal secretory immunoglobulin A is increased in healthy infants who receive a formula with short-chain galactooligosaccharides and long-chain fructooligosaccharides. *The Journal of Nutrition*, 138(6):1141-7.
- Shastri, P., McCarville, J., Kalmokoff, M., Brooks, S. P. J., and Green-Johnson, J. M. (2015). Sex differences in gut fermentation and immune parameters in rats fed an oligofructose-supplemented diet. *Biology of Sex Differences*, 6(13):1-12. Doi: 10.1186/s13293-015-0031-0.
- Vandenplas, Y., Zakharova, I., and Dmitrieva, Y. (2015). Oligosaccharides in infant formula: more evidence to validate the role of prebiotics. *British Journal of Nutrition*, 113:1339-1344.
- Yamamoto, Y., Takahashi, T., To, M., Nakagawa, Y., Hayashi, T., Shimizu, T., Kamata, Y., Saruta, J., and Tsukinoki, K. (2016). The salivary IgA flow rate is increased by high concentrations of short-chain fatty acids in the caecum of rats ingesting fructooligosaccharides. *Nutrients*, 8(5):50-512. Doi: 10.3390/nu8050500.

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